

## REVIEWARTICLE

# Involvement of Regucalcin in Human Carcinogenesis Prevention

**Masayoshi Yamaguchi**

Department of Pathology and Laboratory Medicine, David Geffen School of Medicine, University of California, Los Angeles, USA

## Abstract

Regucalcin, a calcium-binding protein, was discovered in 1978 and is associated with multifunctional role as a suppressor in signal transduction-related translational activity in various types of cells and tissues. Its gene, *rgn*, is located on the X chromosome in humans. Regucalcin also suppresses nuclear deoxyribonucleic acid (DNA) and ribonucleic acid (RNA) synthesis in liver cells. Overexpression of regucalcin suppresses the proliferation of cancer cells by inhibiting G1 and G2/M cell cycle arrest. Regucalcin gene expression is suppressed in cancer tissues of patients, and survival was demonstrated to be prolonged in patients with various types of cancer with higher levels of regucalcin. Suppressed regucalcin gene expression may play a crucial role in the development of carcinogenesis. Development of regucalcin gene delivery system, as a novel gene therapy, is expected to be beneficial in the clinical aspects in the treatment of cancer patients

## Keywords

Regucalcin, cell signaling, nuclear regulation, cell proliferation, carcinogenesis.

## Introduction

Regucalcin was discovered in 1978 as a novel calcium-binding protein that suppresses calcium signaling in various types of cells and tissues.<sup>1-6</sup> The regucalcin gene (gene symbol; *rgn*) is located on X chromosome,<sup>7,8</sup> and organization of the regucalcin gene consists of seven exons and six introns,<sup>9</sup> and it comprises regucalcin family of over 15 species of vertebrate and invertebrate.<sup>5,6,10</sup> Regucalcin plays a multifunctional role in cell regulation: maintains intracellular Ca<sup>2+</sup> homeostasis and suppresses signal transduction, protein synthesis, cell proliferation and apoptosis.<sup>2-4</sup> Regucalcin has been found to play a pivotal role in maintaining cell homeostasis as a suppressor of cell signaling in various types of cells and tissues.<sup>2-4</sup>

Cancer is a pathological condition, where cells display uncontrolled growth, invasion and metastasis. Cell proliferation is mediated through various intracellular signaling transduction-related transcriptional activities that are stimulated by various hormone and cytokines. Enhanced cell proliferation may lead to carcinogenesis. However, mechanism of carcinogenesis is complex and development of novel therapy is required after thorough investigations. Regucalcin has been demonstrated to play a novel suppressing role in cell signaling, and it plays a multifunctional role in regulation of function of various types of cells and tissues.<sup>2-4</sup> Interestingly, overexpression of the regucalcin gene was found to suppress liver cell proliferation and carcinogenesis in animal models.<sup>11-13</sup> Moreover, analysis with multiple gene expression profiles and proteomics has showed that regucalcin gene expression is uniquely suppressed in various human carcinoma tissues.<sup>11-14</sup> Suppressed regucalcin gene expression may lead to the development of carcinogenesis. This mini-review focuses on the potential role of regucalcin as a suppressor in the development of carcinogenesis.



## Open Access

**Citation:** Yamaguchi, M. Involvement of Regucalcin in Human Carcinogenesis Prevention. *Cancer Studies*. 2017; 1(1):2.

Received: June 29, 2017

Accepted: September 10, 2017

Published: September 17, 2017

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**Corresponding author:**

Masayoshi Yamaguchi, Department of Pathology and Laboratory Medicine, David Geffen School of Medicine, University of California, Los Angeles, CA, USA.

E-mail: MasayoshiYamaguchi@mednet.ucla.edu

## Role of Regucalcin as a Suppressor in Cell Proliferation

Regucalcin is localized in the cytoplasm and nucleus in cells.<sup>15,16</sup> Nuclear translocation of cytoplasmic regucalcin has been found to passively transport regucalcin to the nucleus through nuclear pore in cells. Immunocytochemical analysis has showed that regucalcin is localized in the nuclei of the cloned normal rat kidney proximal tubular epithelial NRK52E cells.<sup>17</sup> Nuclear localization of regucalcin is enhanced through hormonal  $Ca^{2+}$ -signaling dependent process, which involves protein kinase C.<sup>17</sup> Regucalcin has been shown to bind protein and DNA and regulate various enzyme activities in the nucleus.<sup>18</sup> Nuclear regucalcin has been found to exhibit suppressive effects on  $Ca^{2+}$ -activated DNA fragmentation by inhibiting endonuclease activity in isolated rat liver nuclei.<sup>19,20</sup> Regucalcin inhibits the activity of Small GTPase Ran (ras-related nuclear protein) that is required for protein export from the nucleus and protein import into the nucleus.<sup>21</sup> Moreover, regucalcin is found to suppress the activities of tyrosine kinase, protein kinase C and  $Ca^{2+}$ /calmodulin-dependent protein kinase, which mediate the process of signal transduction from the cytoplasm to nucleus in cells.<sup>22</sup> In addition, nuclear endogenous regucalcin has been shown to play a suppressive role in the regulation of protein tyrosine phosphatases by using anti-regucalcin monoclonal antibody in the reaction mixture.<sup>23</sup> Thus, regucalcin has been shown to play a crucial role in the regulation of the activity of various enzymes in the nucleus.

Regucalcin has also been demonstrated to exhibit suppressive effects on DNA and RNA synthesis activity in the nuclei of normal rat liver and regenerating rat liver *in vivo*.<sup>24-27</sup> Regucalcin may suppress the enhancement of nuclear DNA and RNA synthesis in proliferating liver cells *in vivo*. Also, regucalcin depressed DNA synthesis activity in the nuclei isolated from rat renal cortex *in vitro*.<sup>28</sup> The presence of anti-regucalcin monoclonal antibody in the reaction mixture containing the liver nucleus causes an increase in nuclear DNA synthesis activity *in vitro*.<sup>24, 25</sup> This increase is completely suppressed in the presence of regucalcin. Thus, endogenous regucalcin is associated with a suppressive effect on DNA synthesis in the nuclei of rat liver and renal cortex.<sup>24,25</sup> The role of regucalcin in inhibiting nuclear RNA synthesis activity in normal rat liver is not observed in the presence of  $\alpha$ -amanitin, an inhibitor of RNA polymerase II and III,<sup>26,27</sup> thereby suggesting its suppressive effect, which is partly resulted from the inhibitory action on RNA polymerase II and III. Regucalcin may be also associated with direct inhibitory effects on nuclear DNA and RNA polymerase activity.

Regucalcin was found to suppress nuclear function in proliferating cells using cloned hepatoma H4-II-E cells cultured in the presence of fetal bovine serum (FBS). Culture with FBS induced an increase in cell number and a corresponding elevation of various kinase activities, which are related to  $Ca^{2+}$ /calmodulin-dependent protein kinase, protein kinase C, protein tyrosine kinase and protein phosphatase activity in H4-II-E cells.<sup>29-31</sup> These enzymes may contribute to the enhancement of hepatoma cell proliferation after serum stimulation. The presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture using H4-II-E cells cultured with FBS stimulation was found to elevate the activities of protein kinase and protein phosphatase. Such an effect is suppressed by the addition of exogenous regucalcin in the enzyme reaction mixture. Regucalcin plays a crucial role as a suppressor in the enhancement of cell proliferation due to inhibiting the activities of various protein kinases and protein phosphatases in the cytoplasm and nucleus.<sup>29-31</sup> Importantly, nuclear DNA synthesis activity was increased at 6 hours after culture with FBS, which is preceded by an elevation in the number of H4-II-E cells cultured with FBS.<sup>32,33</sup> Nuclear DNA synthesis activity in H4-II-E cells was significantly suppressed by the addition of regucalcin in the reaction mixture, and its activity was enhanced by the addition of regucalcin into the reaction mixture,<sup>32, 33</sup> thereby supporting the view that endogenous regucalcin suppresses DNA synthesis activity by which inhibiting protein kinases in the nuclei of proliferating H4-II-E cells using anti-regucalcin monoclonal antibody.<sup>32</sup>

Moreover, to determine the role of endogenous regucalcin in the regulation of nuclear DNA synthesis, regucalcin/pCXN2-transfected cells, where H4-II-E cells overexpress regucalcin stably, were generated.<sup>33</sup> The increase in cell number and DNA synthesis activity in transfectants was suppressed as compared to those of wild- and mock-type; thereby indicating that overexpression of endogenous regucalcin has suppressive effects on cell proliferation.<sup>33</sup> This finding supported the view that the augmentation of endogenous regucalcin has potent suppressive effects on nuclear DNA synthesis activity in proliferating

hepatoma cells. Regucalcin has been proposed to play a suppressive role for the over-proliferation of liver cells.

Overexpression of regucalcin was demonstrated to induce G1 and G2/M phase cell cycle arrest in transfectants (H4-II-E cells).<sup>34</sup> The mRNA expression of *p21*, an inhibitor of cyclin-dependent kinases (cdk), was increased in transfectants, although *cdc2a* and *chk2* (checkpoint-kinase 2) mRNA levels were not significantly altered.<sup>34</sup> Regucalcin may enhance *p21* expression and inhibits G1 progression in H4-II-E cells. Overexpression of endogenous regucalcin has also been shown to suppress proliferation of cloned normal rat kidney proximal tubular epithelial NRK52E cells.<sup>35</sup> Endogenous regucalcin was shown to induce G1 and G2/M phase cell cycle arrest in NRK52E cells.<sup>35</sup> Interestingly, expression of *c-jun* and *chk2* (checkpoint-kinase 2) mRNAs was suppressed in the transfectants of NRK52E cells,<sup>35</sup> and the expression of *c-myc*, *c-fos*, *cdc2* and *p21* mRNAs was not altered in transfectants.<sup>35</sup> Suppressed *c-jun* and *chk2* mRNA expressions may partly contribute to suppression of cell proliferation induced in regucalcin-overexpressing NRK52E cells. In addition, *c-myc*, *c-fos*, *c-jun*, and *Ha-ras* are known as tumor stimulator genes.<sup>36</sup> *p53* and *Rb* are tumor suppressor genes, and *c-src* is oncogene.<sup>37</sup> Expression of *c-myc*, *Ha-ras* or *c-src* mRNAs was suppressed in regucalcin-overexpressing transfectants.<sup>38</sup> Expression of *p53* and *Rb* mRNAs was markedly enhanced in transfectants.<sup>38</sup> Suppressed expression of *c-myc*, *Ha-ras* and *c-src* mRNAs and enhanced expression of *p53* and *Rb* mRNAs in transfectants may lead to retardation of proliferation of hepatoma H4-II-E cells. Also, expression of *p53* mRNA was enhanced in regucalcin-overexpressing transfectants of NRK52E cells, while expression of *c-myc*, *c-fos*, *cdc2* and *p21* mRNAs was not altered in transfectants.<sup>35</sup> Suppressed *c-jun* and *chk2* mRNA expressions and enhanced *p53* mRNA expression may lead to retardation of cell proliferation in NRK52E cells overexpressing regucalcin.

As described above, regucalcin suppressed the effects on cell proliferation by regulating many gene expressions that are related to cell proliferation in hepatoma H4-II-E cells and normal kidney NRK52E cells.<sup>39</sup> Regucalcin can bind DNA and modulate nuclear transcriptional activity.<sup>18</sup> Also, regucalcin can bind to the promoter region of various genes, which suppress stimulator gene expression or stimulate suppressor gene expression in cell proliferation.<sup>39</sup> Overexpression of endogenous regucalcin suppresses cell proliferation.<sup>39</sup> Regucalcin may play a pivotal role as a suppressor for over-proliferation of normal and cancer cells by regulating multi-signaling process related to transcription activity. Interestingly, overexpression of regucalcin was shown to protect apoptotic cell death in normal and cancer cells induced by various signaling stimulating-factors,<sup>40</sup> supporting the view that the regucalcin-suppressed proliferation is not critical for apoptotic cell death.

## Suppressive Role of Regucalcin in the Development of Carcinogenesis

Hepatocellular carcinoma (HCC), the most common primary liver cancer, is one of the most prevalent malignant diseases worldwide, and the third most common causes of cancer-related death.<sup>41-43</sup> HCC originates on a background of cirrhosis, a chronic and diffuse hepatic disease, which results from continuous liver injury and regeneration.<sup>43</sup> Cirrhosis is present in approximately 80%-90% of HCC patients and constitutes the largest single risk factor. In cirrhotic liver, changes in fat metabolism associated with the activation of adipocyte-like pathways are thought to be involved in neoplastic transformation.<sup>43</sup> Increased hepatocyte turnover, inflammation and oxidative DNA damage is implicated in the pathogenesis of various liver diseases, obesity, type 2 diabetes, insulin resistant and nonalcoholic fatty liver disease. The prevalent risk factors for HCC that includes viral infections (hepatitis B and C) and alcohol consumption; further risk factors include tobacco smoking, exposure to aflatoxin B1 and vinyl chloride, diabetes, and genetic disorders, such as hemochromatosis and alpha-1 antitrypsin deficiency are also found to be associated with the development of liver cirrhosis.<sup>44-48</sup>

Hepatocarcinogenesis is a multistep process initiated by external stimuli that lead to genetic changes in hepatocytes or stem cells, resulting in proliferation, apoptosis, dysplasia and neoplasia. The majority of HCC cases are also related to chronic viral infections. Hepatitis B virus (HBV) DNA integrates into the host genome, inducing chromosome instability and insertional mutations that may activate various oncogenes, such as cyclin A.<sup>49-52</sup> Viral proteins, in particular X protein (HBx), act as transactivators that upregulate several oncogenes, such

as *c-myc* and *c-jun*, and transcriptional factors, such as nuclear factor- $\kappa$ B.<sup>59-61</sup> Additionally, HBx activates promoters of genes encoding interleukin-8 (IL-8), tumor necrosis factor (TNF)- $\alpha$ , transforming growth factor (TGF)- $\beta$  and epidermal growth factor receptor (EGFR).<sup>54</sup> HBx can also stimulate several signal transduction pathways, including the JAK/STAT, RAS/RAF/MAPK, and Wnt/ $\beta$ -catenin pathways.<sup>54,55</sup> The contributions of hepatitis C virus (HCV) to hepatocarcinogenesis are mediated through viral proteins, including core, NS3 and NS5A proteins. HCV core protein can promote apoptosis or cell proliferation by interaction with *p53* or upregulation of *Wnt-1* at the transcriptional level.<sup>56</sup>

The prognosis of advanced HCC remains poor in spite of the development of novel therapeutic strategies. Traditional therapies are not effective for HCC and are too toxic for patients with cirrhosis. Currently, transarterial chemoembolization and radioembolization remain the main treatments for intermediate-stage HCC. Improved knowledge of the oncogenic processes and signaling pathways, which regulate tumor cell proliferation, differentiation, angiogenesis, invasion and metastasis, has led to the identification of several potential therapeutic targets that have driven the development of molecular-targeted therapies.<sup>57</sup> An ideal cancer target meets the following criteria: the target is relatively specific for cancer cells (not expressed or expressed at very low levels in normal cells but overexpressed in cancer cells).<sup>67</sup> The target is "drugable" as an enzyme (e.g., a kinase) or cell surface molecule (e.g., a membrane-bound receptor) that can be easily screened for small-molecule inhibitors or targeted by a specific antibody.<sup>57,58</sup> The only systemic therapy available for advanced HCC is based on the multikinase inhibitor sorafenib,<sup>58</sup> which is the most effective therapeutic tool for advanced nonresectable HCC. In the past few years, the use of sorafenib in combination with transarterial chemoembolization has improved survival rates in patients with advanced HCC. New perspectives in cancer treatment have appeared with the advent of microRNAs, a novel class of noncoding small RNAs.<sup>59</sup>

Regucalcin, a suppressor protein in various cell signal transductions,<sup>3,4</sup> has been demonstrated to play a pivotal role in the suppression of hepatocarcinogenesis.<sup>35, 60</sup> As introduced in previous section, overexpression of regucalcin was found to play a role as a suppressor protein in cell proliferation that is mediated through various signaling stimulations in the cloned normal rat kidney proximal tubular epithelial NRK52E cells and the cloned rat hepatoma H4-II-E cells, inducing G1 and G2/M phase cell cycle arrest.<sup>11</sup> Regucalcin has also been demonstrated to exhibit direct inhibitory effects on the activities of various  $Ca^{2+}$ /calmodulin-dependent enzymes, protein kinases and protein phosphatases in the cytoplasm and nuclei, suppressive effects on nuclear DNA and RNA synthesis, depressive effects on the gene expression of *c-myc*, *Ha-ras* and *c-src*, a tumor-stimulator gene and stimulatory effects on the gene expression of *p53* and *Rb*, a tumor-suppressor gene.<sup>39</sup> Moreover, regucalcin was demonstrated to inhibit protein synthesis by inhibiting aminoacyl-tRNA synthetase and stimulate protein degradation by activating cysteinyl protease.<sup>3,4</sup> Thus, suppressive effects of regucalcin on cell proliferation are mediated by targeting multi-molecules in liver cells.

Importantly, the gene expression of regucalcin was demonstrated to be suppressed in the development of hepatocarcinogenesis. Liver regucalcin gene expression was suppressed at early periods of carcinogenesis in rats treated with diethylnitrosamine and then 2-acetylaminofluorene combined with partial hepatectomy, which induces an increase in proliferating cells.<sup>12</sup> Suppression of regucalcin protein expression was identified in proteomic analysis that was differentially expressed in the livers of rats fed 5% ethanol for one and three months,<sup>13</sup> Liver regucalcin mRNA expression was suppressed by liver metabolism disorder induced by administration of carbon tetrachloride,<sup>61</sup> galactosamine<sup>62</sup> and phenobarbital<sup>63</sup> in rats. Hepatic regucalcin level was also reduced in diabetes and during ethanol ingestion,<sup>64</sup> which may result in cirrhosis and HCC. Suppressed regucalcin gene expression may lead to the development of HCC. Multiple gene expression profiles and proteomics analysis have showed that the regucalcin gene and its protein levels were found to be significantly suppressed in human HCC. Suppressed regucalcin gene expression may lead to the development of human hepatocarcinogenesis.

## Prospects

We demonstrated that regucalcin mRNA expression is suppressed in various human normal and tumor tissues, including HCC, kidney transitional cell carcinoma, brain malignant

meningioma and lung non-small cell carcinoma in human subjects.<sup>14</sup> Regucalcin plays a key role in suppressing the cell proliferation and carcinogenesis in various types of human cancer cells and tissues. Importantly, survival has been shown to be prolonged in pancreatic cancer patients, with increased regucalcin gene expression,<sup>70</sup> and overexpression of regucalcin was found to suppress the cell proliferation in human pancreatic cancer MIA PaCa-2 cells *in vitro*.<sup>70</sup> Overexpression of the regucalcin gene in cancer cells may exhibit preventive and therapeutic effects on the development of carcinogenesis. Development of the regucalcin gene deliver system will be expected as a novel gene therapy in clinical aspects for cancer treatment.

## Author Disclosure

The author has no conflicts of interest.

## Acknowledgements

This study was partly supported by the Foundation for Biomedical Research on Regucalcin, Japan.

## References

1. Yamaguchi M, Yamamoto T. Purification of calcium binding substance from soluble fraction of normal rat liver. *Chem Pharm Bull.* 1978;26(6):1915-1918.
2. Yamaguchi M. A novel  $Ca^{2+}$ -binding protein regucalcin and calcium inhibition. Regulatory role in liver cell function. In: Kohama K, ed. *Calcium Inhibition*. Boca Raton, BR: CRC Press; 1992: 19-41.
3. Yamaguchi M. Role of regucalcin in maintaining cell homeostasis and function (Review). *Int J Mol Med.* 2005;15(3): 371-389.
4. Yamaguchi M. Regucalcin and cell regulation: role as a supressor in cell signaling. *Mol Cell Biochem.* 2011;353:101-137.
5. Shimokawa N, Yamaguchi M. Molecular cloning and sequencing of the cDNA coding for a calcium-binding protein regucalcin from rat liver. *FEBS Lett.* 1993;327(3): 251-255.
6. Misawa H, Yamaguchi M. The gene of  $Ca^{2+}$ -binding protein regucalcin is highly conserved in vertebrate species. *Int J Mol Med.* 2000;6(2): 191-196.
7. Shimokawa N, Matsuda Y, Yamaguchi M. Genomic cloning and chromosomal assignment of rat regucalcin gene. *Mol Cell Biochem.* 1995;151(2):157-163.
8. Thiselton DL, McDowall J, Brandau O, et al. An integrated, functionally annotated gene map of the DXS8026-ELK1 internal on human Xp11.3-Xp11.23: Potential hotspot for neurogenetic disorders. *Genomics.* 2002;79(4): 560-572.
9. Yamaguchi M, Makino R, Shimokawa N. The 5' end sequences and exon organization in rat regucalcin gene. *Mol Cell Biochem.* 1996;165(2): 145-156.
10. Yamaguchi M. The transcriptional regulation of regucalcin gene expression. *Mol Cell Biochem.* 2011;346(1-2): 147-171.
11. Yamaguchi M. Suppressive role of regucalcin in liver cell proliferation: Involvement in carcinogenesis. *Cell Prolif.* 2013;46(3): 243-253.
12. Suzuki S, Asamoto M, Tsujimura K, et al. Specific differences in gene expression profile revealed by cDNA microarray analysis of glutathione S-transferase placental form (GST-P) immunohistochemically positive rat liver foci and surrounding tissue. *Carcinogenesis.* 2004;25(3): 439-443.
13. Fernando H, Wiktorowicz JE, Soman KV, et al. Liver proteomics in progressive alcoholic steatosis. *Toxicol Appl Pharmacol.* 2013;266(3): 470-480.
14. Murata T, Yamaguchi M. Alternatively spliced variants of the regucalcin gene in various human normal and tumor tissues. *Int J Mol Med.* 2014;34(4):1141-1146.
15. Tsurusaki Y, Yamaguchi M. Suppressive effect of endogenous regucalcin guanosine triphosphatase activity in rat liver nucleus. *Biol Pharm Bull.* 2001;24(8): 958-961.
16. Tsurusaki Y, Misawa H, Yamaguchi M. Translocation of regucalcin to rat liver nucleus: Involvement of nuclear protein kinase and protein phosphatase regulation. *Int J Mol Med.* 6(6): 655-660.
17. Nakagawa T, Yamaguchi M. Nuclear localization of regucalcin is enhanced in culture with protein kinase

- C activation in cloned normal rat kidney proximal tubular epithelial NRK52E cells. *Int J Mol Med.* 2008;21(5):605-610.
18. Tsurusaki Y, Yamaguchi M. Role of regucalcin in liver nuclear function: Binding of regucalcin to nuclear protein or DNA and modulation of tumor-related gene expression. *Int J Mol Med.* 2004;14(2):277-281.
  19. Jones DP, McConkey DJ, Nicotera P, et al. Calcium-activated DNA fragmentation in rat liver nuclei. *J Biol Chem.* 1989;264(11): 6398-6403.
  20. Yamaguchi M, Sakurai T. Inhibitory effect of calcium-binding protein regucalcin on Ca<sup>2+</sup>-activated DNA fragmentation in rat liver nuclei. *FEBS Lett.* 1991;279(2): 281-284.
  21. Moroiaru J, Blobel G. Protein export from the nucleus requires the GTPase Ran and GTP hydrolysis. *Proc Natl Acad Sci USA.* 1995;92(10): 4318-4322.
  22. Katsumata T, Yamaguchi M. Inhibitory effect of calcium-binding protein regucalcin on protein kinase activity in the nuclei of regenerating rat liver. *J Cell Biochem.* 1998;71(4): 569-576.
  23. Omura M, Yamaguchi M. Regulation of protein phosphatase activity by regucalcin localization in rat liver nuclei. *J Cell Biochem.* 1999;75(3): 437-445.
  24. Yamaguchi M, Kanayama Y. Calcium-binding protein regucalcin inhibits deoxyribonucleic acid synthesis in the nuclei of regenerating rat liver. *Mol Cell Biochem.* 1996;162(2): 121-126.
  25. Tsurusaki Y, Yamaguchi M. Suppressive role of endogenous regucalcin in the enhancement of deoxyribonucleic acid synthesis activity in the nucleus of regenerating rat liver. *J Cell Biochem.* 2002;85(3): 516-552.
  26. Yamaguchi M, Ueoka S. Inhibitory effect of calcium-binding protein regucalcin on ribonucleic acid synthesis in isolated rat liver nuclei. *Mol Cell Biochem.* 1997;173: 169-175.
  27. Tsurusaki Y, Yamaguchi M. Role of endogenous regucalcin in nuclear regulation of regenerating rat liver: Suppression of the enhanced ribonucleic acid synthesis activity. *J Cell Biochem.* 2002;87: 450-457.
  28. Morooka Y, Yamaguchi M. Suppressive effect of endogenous regucalcin on deoxyribonucleic acid synthesis in the nuclei of rat renal cortex. *Mol Cell Biochem.* 2002;229(1-2): 157-162.
  29. Inagaki S, Yamaguchi M. Suppressive role of endogenous regucalcin in the enhancement of protein kinase activity with proliferation of cloned rat hepatoma cells (H4-II-E). *J Cell Biochem.* 2001;36: 12-18.
  30. Inagaki S, Yamaguchi M. Enhancement of protein tyrosine phosphatase activity in the proliferation of cloned rat hepatoma H4-II-E cells: Suppressive role of endogenous regucalcin. *Int J Mol Med.* 2002;6(3): 323-328.
  31. Inagaki S, Misawa H, Yamaguchi M. Role of endogenous regucalcin in protein tyrosine phosphatase regulation in the cloned rat hepatoma cells (H4-II-E). *Mol Cell Biochem.* 2000;213(1-2): 43-50.
  32. Inagaki S, Yamaguchi M (2001) Regulatory role of endogenous regucalcin in the enhancement of nuclear deoxyribonucleic acid synthesis with proliferation of cloned rat hepatoma cells (H4-II-E). *J Cell Biochem.* 2001;82: 704-711.
  33. Misawa H, Inagaki S, Yamaguchi M. Suppression of cell proliferation and deoxyribonucleic acid synthesis in cloned rat hepatoma H4-II-E cells overexpressing regucalcin. *J Cell Biochem.* 2001;84(1): 143-149.
  34. Yamaguchi M, Daimon Y. Overexpression of regucalcin suppresses cell proliferation in cloned rat hepatoma H4-II-E cells: Involvement of intracellular signaling factors and cell cycle-related genes. *J Cell Biochem.* 2005;95(6): 1169-1177.
  35. Nakagawa T, Sawada N, Yamaguchi M. Overexpression of regucalcin suppresses cell proliferation of cloned normal rat kidney proximal tubular epithelial NRK52E cells. *Int J Mol Med.* 2005;16: 637-664.
  36. Curran T. Fos and Jun: Intermediary transcription factors. In: Cohen P, Foulkes JG, ed. The hormonal control of gene transcription, New York, NY: Elsevier Science Publisher; 1991: 295-308.
  37. Hulla JE, Schneider RP. Structure of the rat p53 tumor suppressor gene. *Nucleic Acids Res.* 1993;21(3): 713-717.
  38. Tsurusaki Y, Yamaguchi M. Overexpression of regucalcin modulates tumor-related gene expression in cloned rat hepatoma H4-II-E cells. *J Cell Biochem.* 2003;90(3): 619-626.
  39. Yamaguchi M. Role of regucalcin in cell nuclear regulation: involvement as a transcriptional factor. *Cell Tissue Res* 2013;354(2): 331-342.

40. Yamaguchi M. The anti-apoptotic effect of regucalcin is mediated through multisignaling pathways. *Apoptosis*. 2013;18(10): 1145-1153.
41. Jemal A, Center MM, DeSantis C, et al. Global patterns of cancer incidence and mortality rates and trends. *Cancer Epidemiol Biomarkers Prev* 2005;19(8): 1893-1907.
42. Bosch FX, Ribes J, Diaz M, et al. Primary liver cancer: worldwide incidence and trends. *Gastroenterology*. 2004;127: S5-S16.
43. Bosch FX, Ribes J, Diaz M, et al. Epidemiology of hepatocellular carcinoma. *Clin Liver Dis*. 2005;9: 191-121.
44. Farazi PA, DePinho RA. Hepatocellular carcinoma pathogenesis: from genes to environment. *Nat Rev Cancer*. 2006;6(9): 674-687.
45. Purohit V, Rapaka R, Kwon OS, et al. Roles of alcohol and tobacco exposure in the development of hepatocellular carcinoma. *Life Sci*. 2013;92(1): 3-9.
46. Wu HC, Santella R. The role of aflatoxins in hepatocellular carcinoma. *Hepat Mon*. 2012;12: e7238. doi: 10.5812/hepatmon.7238
47. Starley BQ, Calcagno CJ, Harrison SA. Nonalcoholic fatty liver disease and hepatocellular carcinoma: a weighty connection. *Hepatology*. 2010;51(5): 1820-1832.
48. Dragani TA. Risk of HCC: genetic heterogeneity and complex genetics. *J Hepatol* 2010;52(2): 252-257.
49. Terasaki S, Kaneko S, Kobayashi K, et al. Histological features predicting malignant transformation of nonmalignant hepatocellular nodules: a prospective study. *Gastroenterology*. 1998;115(5): 1216-1222.
50. Brechot C, Pourcel C, Louise A, et al. Presence of integrated hepatitis B virus DNA sequences in cellular DNA of human hepatocellular carcinoma. *Nature* 1980;286(5772): 533-535.
51. Minami M, Daimon Y, Mori K, et al. Hepatitis B virus-related insertional mutagenesis in chronic hepatitis B patients as an early drastic genetic change leading to hepatocarcinogenesis. *Oncogene* 2005;24: 4340-4348.
52. Wang J, Chenivresse X, Henglein B, et al. Hepatitis B virus integration in a cyclin A gene in a hepatocellular carcinoma. *Nature* 1990;343: 555-557.
53. Chirillo P, Falco M, Puri PL, et al. Hepatitis B virus pX activates NF-kappa B dependent transcription through a Raf-independent pathway. *J Virol*. 1996;70: 641-646.
54. Andrisani OM, Barnabas S. The transcriptional function of the hepatitis B virus X protein and its role in hepatocarcinogenesis (Review). *Int J Oncol*. 1999;15(2): 373-379.
55. Cha MY, Kim CM, Park YM, et al. Hepatitis B virus X protein is essential for the activation of Wnt/beta-catenin signaling in hepatoma cells. *Hepatology*. 2004;39(6): 1683-1693.
56. Fukutomi T, Zhou Y, Kawai S, et al. Hepatitis C virus core protein stimulates hepatocyte growth: correlation with upregulation of wnt-1 expression. *Hepatology*. 2005;41(5): 1096-1105.
57. Shin JW, Chung Y-H. Molecular targeted therapy for hepatocellular carcinoma: current and future. *World J Gastroenterol*. 19(37): 6144-6155.
58. Wilhelm SM, Adnane L, Newell P, et al. Preclinical overview of sorafenib, a multikinase inhibitor that targets both Raf and VEGF and PDGF receptor tyrosine kinase signaling. *Mol Cancer Ther*. 2008;7(10): 3129-3140.
59. Callegari E, Elamin BK, Sabbioni S, et al. Role of microRNAs in hepatocellular carcinoma: a clinical perspective. *Onco Targets and Therapy*. 2013;6: 1167-1178.
60. Yamaguchi M. Involvement of regucalcin as a suppressor protein in carcinogenesis Insight into the gene therapy. *J Cancer Res Clin Oncol*, 2015; In press.
61. Isogai M, Shimokawa N, Yamaguchi M. Hepatic calcium-binding protein regucalcin is released into the serum of rats administered orally carbon tetrachloride. *Mol Cell Biochem*. 1994;131(2): 174-179.
62. Isogai M, Oishi K, Yamaguchi M. Serum release of hepatic calcium-binding protein regucalcin by liver injury with galactosamine administration in rats. *Mol Cell Biochem*. 1994;136 (1): 85-90.
63. Isogai M, Oishi K, Shimokawa N, et al. Expression of hepatic calcium-binding protein regucalcin mRNA is decreased by phenobarbital administration in rats. *Mol Cell Biochem*. 1994;141(1): 15-19.
64. Isogai M, Kurota H, Yamaguchi M. Hepatic calcium-binding protein regucalcin concentration is decreased by streptozotocin-diabetic state and ethanol ingestion in rats. *Mol Cell Biochem*.

1997;168(1-2): 67-72.

65. Graveel CR, Jatkoe T, Madore SJ, et al. Expression profiling and identification of novel genes in hepatocellular carcinomas. *Oncogene*. 2001;20(21): 2704-2712.
66. Choia JK, Choi JY, Kim DG, et al. Integrative analysis of multiple gene expression profiles applied to liver cancer study. *FEBS Lett*. 2004;565: 93–100.
67. Blanc J-F, Lalanne C, Plomion C, et al. Proteomic analysis of differentially expressed proteins in hepatocellular carcinoma developed in patients with chronic viral hepatitis C. *Proteomics*. 2005;5(14): 3778–3789.
68. Roy L, LaBoissière S, Abdou E, et al. Proteomic analysis of the transitional endoplasmic reticulum in hepatocellular carcinoma: An organelle perspective on cancer. *Biochim Biophys Acta*. 2010;1804(9): 1869-881.
69. Schröder PC, Segura V, Riezu JI, et al. A signature of six genes highlights defects on cell growth and specific metabolic pathways in murine and human hepatocellular carcinoma. *Funct Integr Genomics*. 2011;11(3): 419–429.
70. Yamaguchi M, Osuka S, Weitzmann MN, et al. Prolonged survival in pancreatic cancer patients with increased regucalcin gene expression: Overexpression of regucalcin suppresses the proliferation in human pancreatic cancer MIA PaCa-2 cells in vitro. *Int J Oncol*. 2016;48(5): 1955-1964.